Biology 113 Closed Book Take-Home Exam #1 – Information Part 1

There is no time limit on this test, though I have tried to design one that you should be able to complete within 3 hours. There are 7 pages in this test, including this cover sheet and the data gallery. You are not allowed to look at someone else’s test, nor use your notes, old tests, the internet, any books, nor are you allowed to discuss the test with anyone until all exams are turned in no later than 12:30 pm on Monday Sept. 24. **EXAMS ARE DUE BY 12:30 pm ON MONDAY SEPTEMBER 24.** If you turn in your exam late, then you lose a letter grade for each day you are late. The **answers to the questions must be typed below each question unless you are instructed to draw something.** If you do not write your answers in the appropriate location, I may not find them.

I have provided you with a “Data Gallery” in the form of figures and tables. To choose a figure in support of your answer, simply state Figure #x. You do NOT need to move the figure on your test. Do not assume how many of the data images you will use, or not use. Simply choosing the data is not sufficient support for your answer. You must explain the significance of the data and how they support your answer. I have given you sentence limits so be concise.

**-3 pts if you do not follow this direction.**

**Please do not write or type your name on any page other than this cover page.**

Staple all your pages (INCLUDING THE TEST PAGES) together when finished with the exam.

Name (please print):

Read the pledge and sign if you can do so with honor:

______________________________

On my honor I have neither given nor received unauthorized information regarding this work, I have followed and will continue to observe all regulations regarding it, and I am unaware of any violation of the Honor Code by others.

How long did this exam take you to complete?
Lab Questions:

6 pts.
1) Below you see three sequences of nucleotides. Label and annotate them as requested.

# line
 ____  ...ACTGCGGCTTAGACTGACATCAGCAGGACGACTGACATGCGGTTTAGA...
 ____  ...TGACCAGGAAGCCTAGCTGAGGACGTGAAGTGCAGGAGCGTGACTGTACGGCAAATCT...
 ____                           AUCAGCAGGACGCUGACAUGCCGUAUUAGA...

a) Use the blanks on the left to label each strand as either DNA or RNA.
b) Draw a box around the sequence or sequences that constitute the promoter.
c) Put a star at the far right end of the strand that served as template for the bottom strand.
d) Label the 5’ and 3’ ends of all three molecules.
e) Draw a circle (that does not look like a box) around the start codon.
f) Number the nucleotides using the standard numbering scheme (use the area to the right of the “# line”). To conserve space, label only every other odd number. To make it clear which nucleotide you are numbering, convert every other odd nucleotide to **bold font**.

4 pts.
2) We are using Golden Gate Assembly to put your new promoters into the receiving plasmid.
a) List two advantages of GGA over traditional cloning methods. Limit each answer to 1 sentence.
   1. 
   2. 

b) How did you figure out what sticky ends to add onto your promoter? Limit each answer to 1 sentence.

Lecture Questions:

6 pts.
2) 

a) In the space below, draw a picture of one ribonucleotide and add the single letter for the base that would reinforce that you have drawn a ribonucleotide. Your drawing should include every atom and bond except those in the base.
b) Number the carbons in your diagram.
c) Add an arrow to show where the next ribonucleotide would be added to the one you have drawn.
6 pts.
3)  
   a) List two common misconceptions about DNA evidence. Limit your answer to 1 sentence for each number.
      1. 
      2. 
   b) When interpreting experimental data, where should you begin your interpretation, and why? Limit your answer to a maximum of 2 sentences.

20 pts.
4)  
   a) Analyze and interpret figure 10 using mathematics to convince a skeptic that the red line indicates a gene was induced rather than protein accumulated at a constant rate simply because cells were replicating. Limit your answer to a maximum of 2 sentences.
   b) In the space below, draw a genetic circuit diagram to explain what was happening at the molecular level when lactose was added to the system in figure 10.
   c) What is the function of LacI and what type of molecule is it? Choose supportive data from the gallery and explain how the data support your answers. Limit your answer to a maximum of 3 sentences.
   d) Compare and contrast steroid regulation of gene activation with lactose gene activation. Choose supportive data from the gallery and explain how the data support your answer. Limit your answer to a maximum of 4 sentences.
   e) Translate this ORF (type your answer in the space below the sequence I supplied). Use the single letter code in your answer.

   GCUAGUCAAUGGCUCUUUGCCUGAUGGUAGCAGACG

9 pts.
5) We have focused on molecular and cellular information. Provide three examples of information that we have studied that are non-linear cell or molecular information. For each example, describe how it illustrates non-linear molecular information. Use this numbered list for your three part answer and limit each number to a maximum of 2 sentences.
10 pts.
6) Describe central dogma. Your answer should be in the format of a **numbered list**. Each line in your outline should be accompanied by supporting data with an explanation of how the data supports your statement. Limit each number to a maximum of 2 sentences.

1…

10 pts.
7) Using a numbered list, compare and contrast three forms of reproduction: bacterial, non-sexual eukaryotic and gamete formation. Provide data for as many of the numbered descriptions as you can. Limit each number to a maximum of 2 sentences.

1…

20 pts.
8) 
   a) Explain why it is evolutionarily advantageous for DNA polymerase to make errors. Limit your answer to a maximum of 2 sentences.
   b) Describe two other sources of variation in sexual reproduction. Support your answer with data and limit each example to 2 sentences.
   c) Use experimental data from the data gallery to explain what happens during the S phase of the cell cycle. Explain how the data support your answer. Limit your answer to a maximum of 3 sentences.
   d) Copy and paste the sequence in question #1 above and indicate three bases that could be modified epigenetically if the DNA came from a human genome. Use an arrow for your visual indication.

   (paste sequence here and delete this line in gray font)

   e) Describe the chemical modification would you make to the bases in part d) above?

9 pts.
9) Bio113 is a magical place and it is not uncommon for students in this class to fall in love and marry after college (love at first sight). Let me share with you some genetic counseling information so you can advise a couple of biology alumni who are planning on having children in the near future. You may use fractions for your answer, and you must show your work to eligible for partial credit.
Bob’s father has the recessive disease abbreviated “XP-F” but he does not. Sue is heterozygous for XP-F.
a) What is the probability of them having a child with XP-F?
b) What is the probability of them having a son with XP-F?
c) What is the probability of them having a son with XP-F or a daughter who is homozygous and disease free?
Data Gallery

1. Infected bacteria
   - Extracellular $^{35}S$
   - Extracellular $^{32}P$

2. Gel images showing bands.
   - Lane 1: Control
   - Lane 2: Experimental

3. Photos showing differences in plant development.
   - a) Normal plant growth
   - b) Mutant plant growth
   - c) Close-up of leaf with abnormal growth

4. UV absorption and RNA yield graph.
   - Small RNAs
   - Large RNAs

5. Graph showing radioactive intensity.
   - Amount of RNA
   - Amount of Radioactivity

6. Gel showing RNA expression.
   - Lane 1: THP
   - Lane 2: TFRB
   - Lane 3: RAP 74
   - Lane 4: RNA pol

7. Frequency distribution of volumes.
   - Volume (μm$^3$)

   - a) Pollen
   - b) Fertilized egg
   - c) Adult plant

9. Gel showing DNA restriction analysis.
   - M: Marker
   - Lanes 1-6: DNA samples

    - Activity (μg)

    - P: Parent
    - X: Cross
    - F1: Generation

12. Graph showing progesterone levels.
    - Incubation Time (min)
    - Cytoplasm
    - Nucleus

13. Radioactivity profile.
    - Aliquot Number
    - Radioactivity (μg)

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Table 3.3 Comparison of fixed independent preparations of the transforming factor and purified DNA.

<table>
<thead>
<tr>
<th>Sample</th>
<th>% carbon, C</th>
<th>% hydrogen, H</th>
<th>% nitrogen, N</th>
<th>% phosphorus, P</th>
<th>NF ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td>31</td>
<td>34.27</td>
<td>3.89</td>
<td>14.23</td>
<td>8.87</td>
<td>1.66</td>
</tr>
<tr>
<td>32</td>
<td>32.78</td>
<td>3.76</td>
<td>13.56</td>
<td>8.04</td>
<td>1.59</td>
</tr>
<tr>
<td>42</td>
<td>32.78</td>
<td>3.76</td>
<td>13.56</td>
<td>8.04</td>
<td>1.59</td>
</tr>
<tr>
<td>44</td>
<td>32.78</td>
<td>3.76</td>
<td>13.56</td>
<td>8.04</td>
<td>1.59</td>
</tr>
<tr>
<td>Purified DNA</td>
<td>34.28</td>
<td>3.81</td>
<td>14.23</td>
<td>8.87</td>
<td>1.66</td>
</tr>
</tbody>
</table>

Position: A: -6.64, C: -6.64, G: -0.37, T: 1.57

Second Base in Codon:

<table>
<thead>
<tr>
<th>UCC</th>
<th>CUC</th>
<th>GCC</th>
<th>CCA</th>
<th>CCU</th>
<th>CCG</th>
<th>GCU</th>
<th>GCA</th>
<th>GCC</th>
<th>GGA</th>
</tr>
</thead>
<tbody>
<tr>
<td>glu</td>
<td>ser</td>
<td>pro</td>
<td>pro</td>
<td>pro</td>
<td>pro</td>
<td>pro</td>
<td>pro</td>
<td>ser</td>
<td>pro</td>
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</table>

First Base in Codon:

<table>
<thead>
<tr>
<th>CUU</th>
<th>CUC</th>
<th>GCU</th>
<th>UGG</th>
<th>UGU</th>
<th>GUG</th>
<th>UUC</th>
<th>UUA</th>
</tr>
</thead>
<tbody>
<tr>
<td>Leu</td>
<td>Leu</td>
<td>Pro</td>
<td>Arg</td>
<td>Arg</td>
<td>Arg</td>
<td>Phe</td>
<td>Phe</td>
</tr>
</tbody>
</table>

Table 2.2 Amount of radioactive RNAs per milligram of total RNA.

<table>
<thead>
<tr>
<th>Type of RNA</th>
<th>Radioactivity after 7 minutes</th>
<th>Radioactivity after 30 minutes</th>
</tr>
</thead>
<tbody>
<tr>
<td>tRNA</td>
<td>6,850</td>
<td>3,850</td>
</tr>
<tr>
<td>tRNA (small)</td>
<td>2,120</td>
<td>2,860</td>
</tr>
<tr>
<td>tRNA (large)</td>
<td>714</td>
<td>2,160</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Genotype</th>
<th>β-galactosidase induction</th>
<th>Permease induction</th>
</tr>
</thead>
<tbody>
<tr>
<td>β' + β'</td>
<td>100</td>
<td>100</td>
</tr>
<tr>
<td>β' + β'</td>
<td>100</td>
<td>100</td>
</tr>
<tr>
<td>β' + β' + β'</td>
<td>1</td>
<td>240</td>
</tr>
<tr>
<td>β' + β'</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>β' + β'</td>
<td>&lt;1</td>
<td>&lt;1</td>
</tr>
</tbody>
</table>

Time Incorporation into long DNA polymers

<table>
<thead>
<tr>
<th>Time</th>
<th>Incorporation into long DNA polymers</th>
</tr>
</thead>
<tbody>
<tr>
<td>0 minutes</td>
<td>14.4</td>
</tr>
<tr>
<td>20 minutes</td>
<td>74.4</td>
</tr>
<tr>
<td>40 minutes</td>
<td>78.6</td>
</tr>
<tr>
<td>80 minutes</td>
<td>82.2</td>
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</table>

<table>
<thead>
<tr>
<th>Generation</th>
<th>Green Peas</th>
<th>Yellow Peas</th>
</tr>
</thead>
<tbody>
<tr>
<td>P</td>
<td>5 true-breeding green plants</td>
<td>5 true-breeding yellow plants</td>
</tr>
<tr>
<td>F1</td>
<td>0 green peas</td>
<td>273 yellow peas</td>
</tr>
<tr>
<td>F1</td>
<td>0 plants from green peas</td>
<td>258 plants from F1, yellow peas</td>
</tr>
<tr>
<td>F2</td>
<td>2,001 green peas</td>
<td>6,022 yellow peas</td>
</tr>
</tbody>
</table>